

A Review on Microsphere with Their Method and Application

Vikas Sharma*¹, Jiyaul Hak², Vatan Chaudhary³, Dr. Prabhakar Vishvakarma⁴

*Correspondence author- Vikas Sharma*¹ (Research Scholar) Department of Pharmacy, IIMT College of medical sciences, IIMT University Meerut 250001, Uttar Pradesh, India*

Jiyaul Hak²-Department of Pharmacy, IIMT College of medical sciences, IIMT University Meerut 250001, Uttar Pradesh, India

Vatan Chaudhary³-Department of Pharmacy, IIMT College of medical sciences, IIMT University Meerut 250001, Uttar Pradesh, India

Dr. Prabhakar Vishvakarma⁴-Department of Pharmacy, IIMT College of medical sciences, IIMT University Meerut 250001, Uttar Pradesh, India

Date of Submission: 01-08-2023

Date of Acceptance: 13-08-2023

Abstract

Microspheres are a kind of multiparticulate drug delivery system that is formulated with the aim of achieving extended or regulated drug release. The primary objectives of using microspheres as drug carriers are to enhance the bioavailability and stability of the medication, as well as to provide targeted administration to a particular place at a predefined pace. Polymeric waxy or other protective compounds, including natural, semi-synthetic, and synthetic polymers, are used in their production. Microspheres are often described as powders with a high degree of flowability, with particle sizes ranging from 1 to 1000 μm . These microspheres are composed of proteins or synthetic polymers. The use of various approaches in the production of microspheres offers a diverse array of possibilities for regulating medication administration parameters and augmenting the therapeutic effectiveness of a certain treatment. Delivery methods such as this provide a multitude of benefits as compared to traditional dose forms. This advantages include enhanced effectiveness, less toxicity, greater patient compliance, and increased convenience. Frequently, these systems use macromolecules as vehicles for the administration of pharmaceutical agents. This paper provides an overview of several kinds of microspheres, various methods of synthesis, their uses, and the evaluation of their efficiency via numerous measures.

KEYWORDS: Microspheres, Types of microspheres, Method of preparation, Application.

I. INTRODUCTION:

Microspheres are solid, spherical particles with a size range of 1-1000 μm . Spherical, freely

moving particles composed of proteins or synthetic polymers are seen. The microspheres are characterized as powders that exhibit free-flowing properties and are composed of proteins or synthetic polymers. These materials possess the ability to undergo biodegradation. There are two distinct classifications of microspheres.

Microcapsules are small spherical structures that consist of a core material surrounded by a shell.

Micromatrices are small-scale arrays or grids composed of microscopic elements.

Microcapsules are characterized by the presence of a clearly defined capsule wall that surrounds the imprisoned material, whereas micromatrices are characterized by the dispersion of the entrapped substance throughout the matrix of microspheres. The use of solid biodegradable microspheres that include a drug distributed or dissolved inside a particle matrix holds promise for achieving controlled drug release. These entities consist of polymeric, waxy, or other substances with protective properties, namely biodegradable manmade polymers and altered natural compounds [1].

Advantages:

Microspheres provide constant and prolonged therapeutic effect.

Reduces the dosing frequency and thereby improve the patient compliance.

They could be injected into the body due to the spherical shape and smaller size.

Better drug utilization will improve the bioavailability and reduce the incidence or intensity of adverse effects [2].

Limitation:

Some of the disadvantages were found to be as follows.

The costs of the materials and processing of the controlled release preparation are substantially higher than those of standard formulations.

The fate of polymer matrix and its effect on the environment

The fate of polymer additives such as plasticizers, stabilizers, antioxidants and fillers

Reproducibility is less.

Process conditions like change in temperature, pH, solvent addition, and evaporation/agitation may influence the stability of core particles to be encapsulated.

The environmental impact of the degradation products of the polymer matrix produced in response to heat, hydrolysis, oxidation, solar radiation or biological agent [3].

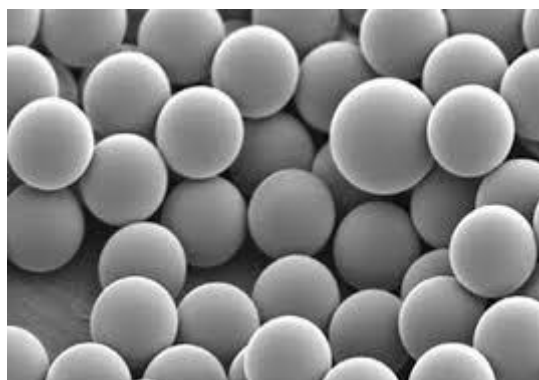


Fig no 1 Structure of Microsphere

CRITERIA FOR MICROSPHERE PREPARATION:

The process of integrating solid, liquid, or gaseous substances into one or several polymeric coverings may be achieved by the use of the microencapsulation technology. [4]

The selection of microsphere production techniques is contingent upon factors such as particle size, route of administration, and length of drug release. These aforementioned characteristics are influenced by variables like rpm, cross-linking method, cross-linking agent, evaporation time, and co-precipitation [5]. The preparation of microspheres must adhere to certain standards.

The ability to incorporate reasonably high concentrations of the drug

- Stability of the preparation after synthesis with a clinically acceptable shelf life
- Controlled particle size and dispersability in aqueous vehicles for injection
- Release of active reagent with a good control over a wide time scale
- Biocompatibility with a controllable biodegradability and
- Susceptibility to chemical modification

TYPES OF MICROSPHERES:

1. Bioadhesive Microspheres:

Adhesion refers to the process by which a medication adheres to a membrane by the use of the adhesive properties inherent in water-soluble polymers. The word "bioadhesion" refers to the attachment of a drug delivery device to mucosal membranes, including buccal, ophthalmic, rectal, nasal, and other relevant sites. These particular microspheres demonstrate an extended duration of stay at the site of administration, allowing close proximity to the absorption site and resulting in enhanced therapeutic efficacy.

2. Magnetic Microspheres:

The use of this specific delivery method has considerable significance due to its ability to facilitate the targeted localization of medications to the precise site of the sickness. The replacement of a smaller amount of magnetically targeted medication has the potential to replace the need for a larger dose of medication that is distributed evenly throughout the body. Magnetic carriers, such as chitosan and dextran, demonstrate magnetic properties when exposed to a magnetic field as a result of the inclusion of magnetic elements in the form of magnetic microspheres. There are several categories.

Therapeutic magnetic microspheres are used for the precise administration of chemotherapeutic drugs to

hepatic neoplasms. This technology also enables the precise delivery of medications, such as proteins and peptides, to specific targets.[6-10]

3. Floating microspheres:

In the context of floating kinds, it is observed that the bulk density is lower than that of gastric fluid, allowing it to stay buoyant inside the stomach without exerting any influence on the pace of gastric emptying. If the drug delivery system is buoyant in stomach content, it may effectively control the release rate of the medication, leading to prolonged gastric residency and increased variability in plasma concentration. Additionally, it also decreases the likelihood of encountering striking incidents and dosage dumping. Another mechanism by which it generates a sustained therapeutic impact is by the reduction of dose frequencies. The administration of the drug ketoprofen is facilitated with this particular formulation.

4. Radioactive microspheres:

Radioembolization treatment involves the use of microspheres with a size range of 10-30 nm, which are bigger than capillaries. These microspheres get trapped in the first capillary bed they encounter. The substances are administered through injection into the arteries that provide blood to the specific tumor under investigation. In each of these scenarios, radioactive microspheres effectively provide a substantial radiation dosage to specific regions, while minimizing harm to adjacent healthy tissues [11]. The radioactivity of microspheres distinguishes them from medication delivery systems, since they do not discharge radioactivity but instead exert their effects within a typical distance associated with a certain radioisotope. These radioactive microspheres may be classified into many types, including α emitters, β emitters, and γ emitters.

5. Polymeric microspheres:

Polymeric microspheres may be categorized into many categories.

i) Polymeric microspheres that is biodegradable

Natural polymers, such as starch, are used because to their inherent characteristics of biodegradability, biocompatibility, and bioadhesiveness. The high degree of swelling property of biodegradable polymers in aqueous medium leads to the creation of a gel, hence prolonging the residence period upon contact with mucosal membranes. The concentration of the polymer directly influences both the pace and amount of drug release, hence regulating the sustained release pattern. One of the primary limitations in the therapeutic use of biodegradable microspheres pertains to the intricate nature of drug

loading efficiency, which poses challenges in effectively regulating the release of drugs. Nevertheless, microspheres provide a diverse.

ii) Synthetic polymeric microspheres

Synthetic polymeric microspheres have garnered significant attention in clinical applications due to their versatile use as bulking agents, fillers, embolic particles, and drug delivery vehicles. Furthermore, extensive research has shown their safety and biocompatibility. However, a significant drawback associated with these types of microspheres is their propensity to migrate from the injection site, hence posing possible risks such as embolism and subsequent organ damage.

METHOD OF PREPARATION:

Spray Drying Technique:

Polymeric blended microspheres loaded with the ketoprofen medication were produced using this method. The process entails the dispersion of the central material into a liquid coating substance, which is then sprayed into the surrounding environment to facilitate the solidification of the coating. This is then followed by the quick evaporation of the solvent [15]. Drug-loaded microspheres were produced by preparing organic solutions of poly(epsilon-caprolactone) (PCL) and cellulose acetate butyrate (CAB) in various weight ratios, together with ketoprofen. These solutions were then sprayed under varied experimental conditions. The quick drying process may result in a loss of crystallinity.

Solvent Evaporation:

The aforementioned procedure is conducted inside a liquid-based production medium. The dispersion of the microcapsule coating is achieved by using a volatile solvent that is not capable of mixing with the liquid production vehicle phase. The core material is dissolved or disseminated inside the covering polymer solution for microencapsulation. The core material combination is distributed in the liquid production vehicle phase with agitation in order to achieve the desired size of microcapsules. Subsequently, if required, the mixture is subjected to heating in order to facilitate the evaporation of the solvent. This process enables the dispersion of the polymer of the core material within the polymer solution, leading to the subsequent contraction of the polymer around the core. Matrix-type microcapsules are generated when the core material is dissolved in the coated polymer solution. The core components might exist in two forms: water-soluble or water-insoluble compounds. Solvent evaporation

entails the creation of an emulsion between a polymer solution and a continuous phase that is immiscible.

Single emulsion technique:

The micro particle carriers composed of natural polymers, namely proteins and carbohydrates, are fabricated using the single emulsion approach. The natural polymers are dissolved or dispersed in an aqueous media and then disseminated in a non-aqueous medium, such as oil. The subsequent phase involves the implementation of cross-linking for the scattered globule. Crosslinking may be accomplished by the use of heat or through the utilization of chemical crosslinking agents. The chemical crosslinking agents used in this study include glutaraldehyde, formaldehyde, and acid chlorides, among others. The process of heat denaturation is not appropriate for compounds that are susceptible to degradation. One drawback of chemical cross-linking is the potential for increased exposure of the active component to chemicals when it is introduced during preparation and then exposed to centrifugation, washing, and separation processes. The characteristics of the surfactants used for the purpose of stabilizing the emulsion phase may significantly impact several aspects of the final multi-particulate product, including its size, size distribution, surface morphology, loading capacity, drug release profile, and bio performance.[12-16]

Double emulsion technique:

The manufacture of microspheres using the double emulsion technique entails the creation of several emulsions or double emulsions of the w/o/w type. This process is particularly well-suited for the encapsulation of water-soluble medicines, peptides, proteins, and vaccines. This methodology is applicable to both natural and manmade polymers. The protein solution, which is in an aqueous form, is disseminated inside a continuous phase composed of lipophilic chemical compounds. The protein solution potentially contains the bioactive components. The continuous phase typically comprises a polymer solution that ultimately encapsulates the protein present in the scattered aqueous phase. The initial emulsion undergoes homogenization or sonication before to being added to the aqueous solution of poly vinyl alcohol (PVA). Consequently, the outcome is the creation of a double emulsion. Subsequently, the emulsion undergoes solvent removal using either solvent evaporation or solvent extraction. The process of double emulsion solvent evaporation/extraction has been effectively used to include various hydrophilic pharmaceuticals, such as luteinizing hormone

releasing hormone (LH-RH) agonist, vaccines, proteins/peptides, and conventional compounds, into microspheres.

Coacervation Method:

The coacervation thermal change experiment included the dissolution of a measured quantity of ethyl cellulose in cyclohexane with the application of vigorous agitation at a temperature of 80°C. Subsequently, the medication underwent a meticulous pulverization process before being introduced into the aforementioned solution. The addition was accompanied by vigorous stirring. To induce phase separation, the temperature was reduced and an ice bath was used. The aforementioned product underwent two cycles of washing with cyclohexane and subsequent air drying. It was then subjected to sieving using a No. 40 sieve to get individual microcapsules. The coacervation non-solvent addition method was used by dissolving a predetermined quantity of ethyl cellulose in toluene, which was supplemented with propylisobutylene, in a sealed beaker. The mixture was subjected to magnetic stirring at a rate of 500 revolutions per minute for a duration of 6 hours. Subsequently, the medication was disseminated throughout the solution, and stirring was maintained for an additional 15 minutes. Phase separation is conducted using petroleum benzoin, with a total of five repetitions and constant stirring. Following the aforementioned steps, the microcapsules underwent a thorough washing process using n-hexane, followed by air drying for duration of 2 hours. Subsequently, the microcapsules were subjected to further drying in an oven set at a temperature of 50 degrees Celsius for a period of 4 hours.

Spray drying and spray congealing:

The techniques used in these systems rely on the evaporation of the polymer and drug solution into a mist inside the surrounding air. The two procedures, namely spray drying and spray congealing, are termed based on the removal of the solvent or chilling of the solution, respectively. The polymer is first dissolved in an appropriate volatile organic solvent, such as dichloromethane or acetone. The solid medication is then disseminated throughout the polymer solution by the process of high-speed homogenization. Subsequently, the dispersion undergoes atomization with the introduction of a high-temperature air stream. Atomization results in the generation of microscopic droplets or fine mist, from which the solvent rapidly evaporates. This process facilitates the creation of microspheres within a size range of 1-100 µm. The process of separating micro particles from hot air is achieved

by the use of a cyclone separator, which effectively isolates these particles. Additionally, the residual solvent present in the air is eliminated through the application of vacuum drying techniques. One of the primary benefits of this method is its ability to be conducted in aseptic conditions, hence ensuring practicality. The spray drying technique is used for the encapsulation of various forms of penicillin. The process of spray congealing is used to encapsulate thiamine mononitrate and sulphathiazole inside a combination of mono- and diglycerides derived from stearic acid and palmitic acid. The fast evaporation of solvents results in the creation of micro particles with a porous structure.

Solvent extraction

The manufacture of micro particles often uses the solvent evaporation process, which entails the extraction of the non-aqueous solvent to remove the organic phase. This approach utilizes water-miscible organic solvents such as isopropanol. The organic phase may be separated using the process of aqueous extraction. This method reduces the duration required for the hardening of the microspheres. One alternative method is the direct integration of the medication or protein into a polymer organic solution. The rate at which solvent is extracted by the extraction process is contingent upon many factors, including the temperature of the water, the ratio of emulsion volume to water, and the solubility profile of the polymer.[17-19]

Quasi Emulsion Solvent Diffusion:

The literature has documented a unique approach using quasi-emulsion solvent diffusion for the production of controlled release drug microspheres using acrylic polymers. Microspheres may be produced by a quasi-emulsion solvent diffusion technique, wherein an external phase comprising distilled water and polyvinyl alcohol is used. In order to increase plasticity, a mixture of drug, ethanol, and polymer is incorporated into the internal phase at a concentration of 20% relative to the polymer content. Initially, the internal phase is produced at a temperature of 60°C and afterwards combined with the exterior phase at ambient temperature. Following the emulsification process, the mixture undergoes continuous stirring for duration of 2 hours. Subsequently, the mixture might undergo filtration in order to achieve the separation of the micro sponges. Subsequently, the product undergoes a washing and drying process using a vacuum oven set at a temperature of 40°C for duration of one day.

FACTORS AFFECTING PARTICLE SIZE, ENTRAPMENT EFFICIENCY AND RELEASE CHARACTERISTICS:

The drug release is strongly influenced by a various parameters including the drug content, the nature of polymer, the physical state of the drug, the molecular weight of polymer, the density of cross linking the copolymer concentration, the type of any excipients included in the micro particles preparation, and the microsphere size.

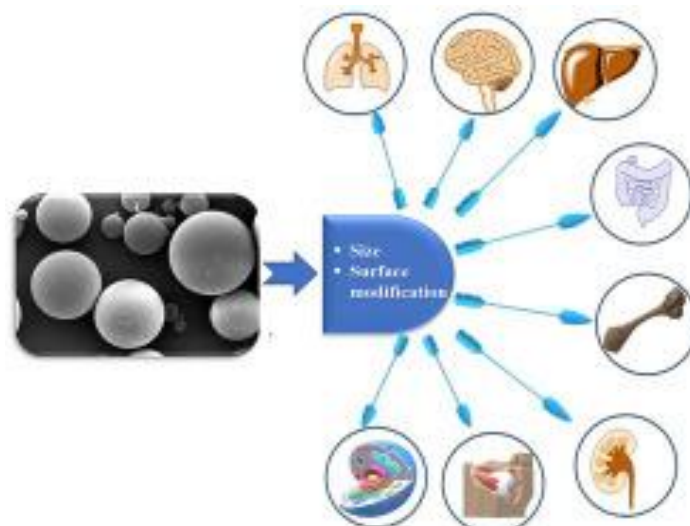


Fig no 2 microsphere efficiency

i) Drug content:

The amount of drug that present in the micro particle determines the release kinetics of the drugs from the matrix devices; the release proportionately increases with increase in drug content in the micro particles.

ii) Nature of polymer:

The nature of polymer present in micro particles and the type of polymer erosion clearly determine the drug delivery rate. Polymers are generally classified into two types: surface erosion and bulk-erosion. In bulk-eroding polymers, the matrix degrades by diffusion of water molecules. While, in surface-leaching polymers, water repelling monomers resist penetration of water molecules therefore degradation takes place from the surface of the particle.

iii) Physical state of the drug:

The physical state of a drug affects the drug release kinetics from a dosage form. The presence of the drug inside the micro particles may vary from molecular dispersion to well defined crystalline structures.

iv) Molecular weight of polymer:

Molecular weight of polymer plays a major role in polymer degradation as well as drug delivery rates. This indicates that, higher the molecular weight lower the diffusivity and decreased drug delivery rate. In addition, drug delivery takes place by diffusion through water filled pore. The decrease in delivery rates reported for small molecules such as drugs, and macromolecules with increasing molecular weight of polymer.[20-25]

v) Density of cross linking:

The cross linking density plays a major role on the release kinetics of drugs from the micro particles. It was observed from the results that drug delivery rates become slower when micro particles preparation utilizes polymer at higher concentration and polymer with higher molecular weight and/or a lower drug concentration.

vi) Co-polymer concentration:

The concentration of co-monomer presence in copolymers has a strong effect on release rates. Normally, the release rate increases with increasing the concentration of polymer that degrades faster. Likewise, when polymer erosion controls the drug delivery, release rate is usually increased by higher concentration of more soluble and/or the smaller monomer.

vii) Type of excipients:

To maintain stability of the drug a range of excipients might be included to micro particle preparations during manufacture and/or release. Decreased delivery rate may be due to interaction with the excipients and forming chelation, complexation, polymerization, isomerisation, racemisation etc.

Evaluation of Microspheres:

Particle size analyzer:

Microspheres weighing 50 mg are suspended in 5 mL of distilled water containing a 2% w/v concentration of tween 80. This is done to avoid aggregation of the microspheres. The suspension is then subjected to sonication in a water bath, and the resulting particle size is measured and reported as the volume mean diameter in micrometers.

Optical microscopy:

The particle size is determined by the use of an optical microscope, namely the Meizer OPTIK. The measurement was conducted with a magnification of 450x (10x eyepiece and 45x objective), and a total of 100 particles were quantified.

Scanning electron microscopy (SEM):

The technique of scanning electron microscopy (SEM) is used to determine the surface morphology. The microcapsules were affixed to the SEM sample slab using double-sided adhesive tape and then coated with a layer of gold film in a vacuum environment. These samples were then subjected to analysis. [26-30]

Swelling index:

The swelling index is a method used to assess the properties of sodium alginate microspheres. Various solutions (100mL) were used, including distilled water, buffer solutions with pH values of 1.2, 4.5, and 7.4, as well as alginate microspheres (100mg). These substances were put in a wire basket and positioned on top of the aforementioned solution. Subsequently, the samples were permitted to undergo swelling at a temperature of 37°C. The measurement of weight fluctuation resulting from swelling in microspheres involves regularly weighing the microspheres after soaking them with filter paper.

Entrapment efficiency:

The measure of the way a system or method captures or contains a desired substance or entity. The microspheres, which encapsulate a medication with a concentration of 5mg, undergo crushing followed by dissolution in distilled water using an ultrasonic stirrer for a duration of 3 hours. The

resulting solution is then filtered and subjected to analysis using UV-Vis spectroscopy. The entrapment efficiency may be defined as the ratio between the actual drug content and the theoretical drug content.

X-ray diffraction:

X-ray diffraction is a scientific technique that involves the interaction of X-rays with a crystalline material, resulting in the scattering of approach described herein may be used to ascertain changes in the crystallinity of a pharmaceutical compound. The analysis of micro particles and their distinct components is facilitated by the use of an X-ray diffraction (XRD) instrument. Scanning range angle between 80°C - 70°C

Thermal analysis:

Thermal analysis is a scientific method used to study the behavior of materials under different temperature conditions. The thermal examination of microcapsules and their components may be conducted via the use of two techniques: Differential Scanning Calorimetry (DSC) and Thermo Gravimetric examination (TGA). Differential thermometric analysis (DTA) is a thermal analysis technique that is often used in scientific research and industrial applications. The sample is precisely weighed and subjected to heating on an alumina pan at a consistent rate of 10°C per minute, while being exposed to a nitrogen flow of 40 ml per minute.

FTTR:

The determination of drug-polymer interaction and drug degradation during the microencapsulation process may be achieved using Fourier Transform Infrared Spectroscopy (FTIR).[31-33]

Stability studies

The topic of investigation pertains to stability research. Stability studies are conducted by putting the microspheres into screw-capped glass containers and subjecting them to storage under certain conditions: The prevailing atmospheric moisture level. The ambient temperature inside the room was maintained at an average of 27 degrees Celsius, with a standard deviation of 2°C.

- The oven temperature is maintained at 40±2°C.
- The refrigerator operates at a temperature of 50±8 oC.
- The experiment was conducted over a period of 60 days, during which the drug content of the microsphere was analyzed [34].

The concept of zeta potential is a crucial parameter in the field of colloid science and surface

chemistry. The polyelectrolyte shell is formed by the inclusion of chitosan with varying molecular weights into the W2 phase. The resultant particles are then analyzed using zeta potential testing.

1. The use of microspheres in many fields and industries

The process of introducing genetic material into a target cell or organism, often referred to as gene delivery, is a fundamental technique in molecular biology and genetic engineering.

2. The Delivery of Drugs to the Ocular System

- Intratumoral and local medication delivery refers to the targeted administration of therapeutic agents directly into the tumor or its immediate vicinity.
- The topic of discussion pertains to the administration of drugs via the oral route.
- Nasal drug delivery is a method of administering medications via the nasal route.
- Buccal drug delivery refers to the administration of pharmaceutical substances via the buccal mucosa, which is the lining of the inner cheek.
- The topic of interest is gastrointestinal medication delivery.
- Peroral drug delivery refers to the administration of medications via the oral route, often known as swallowing.
- Vaginal medication delivery refers to the administration of pharmaceutical substances via the vaginal route.
- Transdermal drug delivery refers to the administration of pharmaceutical substances via the skin, allowing for systemic absorption into the bloodstream.
- Drug distribution to the colon

The multiparticulate delivery system is a method used for the administration of drugs or other therapeutic agents in a multiparticulate form.

II. CONCLUSION:

The concept of "microsphere" is often used to refer to small spherical particles, and it has found extensive use in the field of drug delivery systems. The primary focus is on targeted drug delivery methods, such as the use of bioadhesive microspheres for nasal, ocular, buccal, rectal, and other applications. Additionally, magnetic microspheres and radioactive microspheres are employed specifically for tumor treatment. Another significant area of interest is controlled and sustained drug delivery, which involves the use of polymeric microspheres and floating microspheres. The integration of diverse methodologies will lead

to the prominent use of microspheres in innovative drug delivery systems, with a special focus on applications such as cell sorting, diagnostics, and genetic engineering. The research provides evidence that Microspheres serve as effective carriers for the innovative medication delivery technology.[35]

REFERENCES

- [1]. Chaudhari A, Jadhav K. R, Kadam VJ. An Overview: Microspheres as a Nasal Drug Delivery System. *Int. J. of Pharmaceutical Sciences Review and Res.* 2010; 5.
- [2]. Vyas SP, Khar RK. Targeted and Controlled drug delivery; 7th Edition; Vallabh Prakashan, New Delhi India, 420-445.
- [3]. Sree Giri Prasad B., Gupta V. R. M., Devanna N., Jayasurya K., Microspheres as drug delivery system – A review, *JGTPS.* 2014; 5(3): 1961 -72.
- [4]. Ghulam M., Mahmood A., Naveed A., Fatima R.A., Comparative study of various microencapsulation techniques. Effect of polymer viscosity on microcapsule characteristics, *Pak. J. Sci.* 22 (3), 2009, 291-300.
- [5]. Li, S.P., Kowalski C.R., Feld K.M., Grim W.M., Recent Advances in Microencapsulation Technology and Equipment, *Drug DevInd Pharm.* 14, 1988, 353-376.
- [6]. Alagusundaram. M, MadhuSudana Chetty. C, Umashankari. K, Attuluri Venkata Badarinath, Lavanya. C and Ramkanth. S. Microspheres as a novel drug delivery system – A Review. *International Journal of Chem Tech Research.* 1(3), 2009, 526-534.
- [7]. Patel JK, Patel RP, Amin AF, Patel MM, 4(6).
- [8]. Li S.P, Kowalski C.R, Feld K.M, Grim W.M. 1988. Recent Advances in Microencapsulation Technology and Equipment, *Drug Dev, Ind Pharm.* 14: 353-376.
- [9]. Shanthi N.C, Gupta R, Mahato K.A. 2010 Traditional and Emerging Applications of Microspheres: A Review, *International Journal of Pharm Tech Research;* 2(1):675-681.
- [10]. Najmuddin M, Ahmed A, Shelar S, Patel V, Khan T. 2010. Floating Microspheres Of Ketoprofen: Formulation and Evaluation, *International Journal Of Pharmacy and Pharmaceutical sciences.* 2(2):83-87.
- [11]. Hafeli U, 2002, *Physics and Chemistry Basic of Biotechnology.* Focus on biotechnology. Review. *Radioactive Microspheres for Medical Application,* 7:213-248.
- [12]. Yadav AV, Mote HH. 2008. Development of Biodegradable Starch Microspheres for Intranasal Delivery, *Indian Journal of pharmaceutical Sciences.* 70 (2):170-174.
- [13]. Saralidze K, Leo H, Koole, Menno L, Knetsch W. 2010. Polymeric Microspheres for Medical Applicatio, *Materials;* 3:3357-3564.
- [14]. Trivedi P, Verma L, Garud N. 2008. Preparation and Characterization of Acclofenac Microspheres, *Asian Journal of pharmaceuticals.* 2(2): 110-115.
- [15]. Mathew Sam T., Devi Gayathri S., Prasanth V.V., Vinod B; NSAIDs as microspheres, *The Internet Journal of Pharmacology.* 6(1), 2008, 67 – 73.
- [16]. Ramteke K.H., Jadhav V.B., Dhole S.N., Microspheres: As carrieres used for novel drug delivery system, *IOSRPHR.* 2012; 2(4):44-48.
- [17]. Patel B., Modi V., Patel K., Patel M., Preparation and evaluation of ethyl cellulose microspheres prepared by emulsification emulsification - solvent evaporation method, *International Journal For Research In Management And Pharmacy.* 2012; 1(1):83-91.
- [18]. Patel N. R., Patel D. A., Bharadia P.D., Pandya V., Modi D., Microsphere as a novel drug delivery, *Int. j. pharm. life sci.* 2011;2(8):992-7
- [19]. Ghulam M., Mahmood A., Naveed A., Fatima R.A., Comparative study of various microencapsulation techniques. Effect of polymer viscosity on microcapsule charecterestics, *J. Sci.* 22 (3), 2009, 291-300.
- [20]. Bansal H., kaur S. P., Gupta A. K., Microsphere: Methods of preparation and applications; a comparative study, *Int J Pharm Sci Rev Res.* 2011; 10(1):69-78.
- [21]. Alagusundaram M., Chetty. C. M. S., Umashankari. K, Badarinath A. V., Lavanya. C., Ramkanth. S., Microspheres as a novel drug delivery system - A review, *Int J Chem Tech Res.* 2009;1(3):526-34.
- [22]. Patel N. R., Patel D. A., Bharadia P.D., Pandya V., Modi D., Microsphere as a novel drug delivery, *Int. j. pharm. life sci.* 2011;2(8):992-7.

- [23]. Bansal H., kaur S. P., Gupta A. K., Microsphere: Methods of preparation and applications; a comparative study, *Int J Pharm Sci Rev Res.* 2011; 10(1):69-78.
- [24]. Alagusundaram M., Chetty. C. M. S., Umashankari. K, Badarinath A. V., Lavanya. C., Ramkanth. S., Microspheres as a novel drug delivery system- A review, *Int J Chem Tech Res.* 2009;1(3):526-34.
- [25]. Davis S.S. and Illum L. (1989). Microspheres as drug carrier in drug carrier system, F.H Roerdink and A.M. Kron (Eds), John Wiley and sons Ltd., 1-6
- [26]. Mathew Sam T, Devi Gayathri S, Prasanthv VV, Vinod B. NSAIDs as microspheres, *The Internet Journal of Pharmacology.* 2008; 6: 11-15.
- [27]. Pradesh TS, Sunny M., Varma KH, Ramesh P. Preparation of micro structured hydroxy apatite microspheres using oil in water emulsion, *Bull Matter. Sci.* 2005; 28(5):383-90.
- [28]. Kannan. K., Karar. K.P., Manavalan. R., Formulation and Evaluation of Sustained Release Microspheres of Acetazolamide by Solvent Evaporation Technique, *J. Pharm. Sci& Res.* 2009; 1 (1):36-39.
- [29]. Shaji J., Poddar A., Iyer S., Brain-Targeted Nasal Clonazepam Microspheres, *Indian Journal of pharmaceutical Sciences.* 2009; 71(6): 715-718.
- [30]. Chowdary K.P.R., Suri B.J., Permeability of Ethylene Vinyl Acetate Copolymer Microcapsules: Effect of Solvents, *Indian Journal of pharmaceutical Sciences.* 2003; 65(1):62-66.
- [31]. Soni L.M., Kumar M., Namdeo P.K., Sodium alginate Microspheres for extending drug release: formulation and in vitro evaluation, *International Journal of Drug Delivery.* 2010; 2(1):64-68.
- [32]. Ghulam M., Mahmood A., Naveed A., Fatima R.A., Comparative study of various microencapsulation techniques. Effect of polymer viscosity on microcapsule characteristics, *Pak.J.Sci.* 2009; 22 (3):291-300.
- [33]. Surini S., Anggriani V., Anwar E., Study of Mucoadhesive Microspheres Based on Pregelatinized Cassava Starch Succinate as a New Carrier for Drug Delivery, *J. Med. Sci.* 2009; 9(6):249-256
- [34]. Tamizharsi S., Rathi C.J., Rathi. Formulation and Evaluation of Pentoxifylline-Loaded Poly (ϵ -caprolactone) Microspheres, *Indian Journal of pharmaceutical Sciences.* 2008; 70(3):333-337.
- [35]. Fischer S., Foreg C., Merkle P.H., Gander B., Chitosan Coated Plga-Microspheres-A Modular System for Targeting Drug Delivery, *European Cells and Materials.* 2004; 7:11-12